

## New Eudesmane and Eremophilane Derivatives from *Laggera Alata*

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**Abstract:** From the aerial part of *Laggera alata*, a novel eremophilanoid (**1**) as well as two new eudesmanoids (**2-3**) were isolated. Their structures were elucidated by 2D-NMR technique and X-ray diffraction studies. The cytotoxic activities of these sesquiterpenes were also investigated.

**Keywords:** *Laggera alata*, sesquiterpene, eremophilanoid, eudesmanoid, X-ray diffraction, cytotoxicity.

*Laggera pterodonta* and *Laggera alata* are the only two species of *Laggera* genus found in China. Both of them are used as traditional herbal medicines in southwestern China. Previous investigations of *L. pterodonta* have led to the isolation of 20 new eudesmane derivatives including some cytotoxic ones<sup>1,2</sup>. These interesting findings have prompted us to a phytochemical examination of *L. alata*. Three new compounds, along with 14 known compounds were isolated from the aerial part of the title plant.

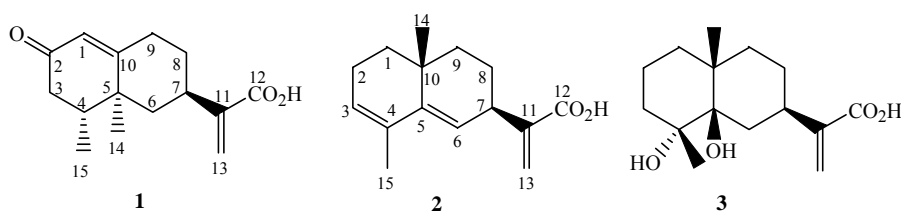
Compound **1** was isolated as colorless needles,  $[\alpha]_D^{25}$  - 83.3 (*c* 0.28, MeOH). Its HREIMS exhibited a  $[M]^+$  at *m/z* 248.141 (calcd. 248.1412) corresponding to a molecular formula C<sub>15</sub>H<sub>20</sub>O<sub>3</sub>. Its IR spectrum (KBr) revealed the presence of an allylic acid moiety (1707 cm<sup>-1</sup>)<sup>3</sup>. <sup>13</sup>C-NMR indicated that it should contain a  $\alpha,\beta$ -unsaturated ketone ( $\delta$  201.9, 146.3, 126.0). <sup>1</sup>H-NMR featured it as an eremophilanoid compound<sup>7</sup>: Me-14 ( $\delta$  1.16, s, 3H), Me-15 ( $\delta$  1.05, d, 3H, *J*=6.0 Hz). Apart from the exomethylene signals observed at  $\delta$  6.26 (br s, 1H) and 5.70 ppm (br s, 1H), another singlet appeared at  $\delta$  5.91 suggesting the presence of a trisubstituted olefin conjugated to a carbonyl group. Based on the above information, the presence of a 1(10)-en-2-one moiety in **1** was deduced. HMBC also revealed that the ketone carbonyl was on C-2 and the olefin carbons were on C-1 and C-10. The stereochemistry of H-7 was presumed to be axial from the coupling constant ( $\delta$  2.64, dddd, 1H, *J*=12.0, 9.0, 4.5, 4.5 Hz). The

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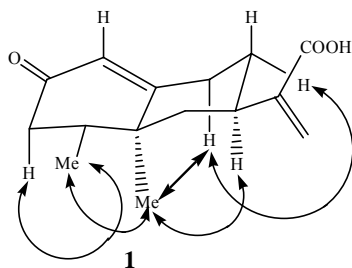
stereochemistry of Me-14 and Me-15 were deduced from the NOESY spectrum, from which clear correlations between H-14 and H-15; between H-14 and H-7 $\alpha$ ; as well as those between H-3 $\alpha$  and H-15 could be observed (**Figure 2**). This was further supported by the CD spectrum that showed a positive Cotton effect at 245 nm and a negative one at 332 nm<sup>4</sup>. The structure of **1** was finally ascertained by X-ray diffraction analysis (**Figure 3**).

**Figure 1** The structures of compounds **1-3**



Compound **2** was obtained as a colorless gum,  $[\alpha]_D^{25} + 7.5$  ( $c$  0.4, CHCl<sub>3</sub>). Its molecular formula was determined as C<sub>15</sub>H<sub>20</sub>O<sub>2</sub> by the fact that the HREIMS exhibited a  $[M]^+$  at  $m/z$  232.1459 (calcd. 232.1463). The <sup>1</sup>H and <sup>13</sup>C-NMR spectra of **2** showed close similarity to those of 11-cinnamoyloxy-3,5-dien-eudesmane (**2a**)<sup>5</sup>. However, the tertiary methyl signals of Me-12 and Me-13 in **2a** were absent in the <sup>1</sup>H and <sup>13</sup>C-NMR spectra of **2**. Instead, signals of a methylene group was observed at  $\delta$  5.72 and 6.34 ppm, suggesting that the isopropyl group in **2a** was replaced by an allylic acid moiety in **2**. This was supported by the IR absorption band of **2** at 1693 cm<sup>-1</sup>, and was further verified by the correlation peaks appearing at the 2D HMQC and HMBC experiments. Since no correlation between H-14 and H-7 was observed in the 2D NOESY spectrum of **2**, the configuration of H-7 should be of the  $\alpha$ -orientation. Therefore, compound **2** was identified as 3,5,11(13)-trien-eudesma-12-oic acid.

**Figure 2** Selective 2D-NOESY correlations of compounds **1**

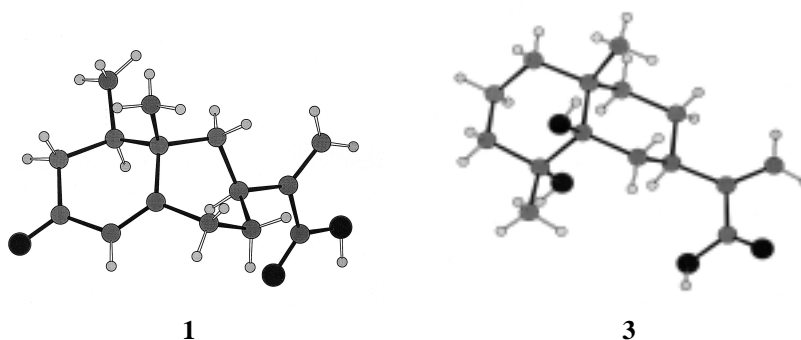


Compound **3** was isolated as colorless needles,  $[\alpha]_D^{25} + 5.39$  ( $c$  0.15, MeOH). The <sup>1</sup>H and <sup>13</sup>C-NMR spectra of **3** bore close resemblance to those of ilicic acid<sup>6,7</sup>. However, the methine carbon signal of C-5 ( $\delta$  55.8) in ilicic acid did not appear in the <sup>13</sup>C-NMR spectrum of **3**. Instead, an oxygenated quaternary carbon resonance exhibited at  $\delta$  76.8. In addition, the C-4 and C-6 of **3** were downfield shifted when comparing with those of

ilicic acid<sup>6,7</sup>. These indicated that **3** was a 5-OH derivative of ilicic acid, consistent with the presence of six methylene signals in the DEPT spectrum of **3**. There was no correlation between H-14 and H-15 in the NOESY spectrum of **3**, suggesting that the A/B ring in the molecular structure of **3** was *cis*-fused. The stereochemistry of H-7 was presumed to be axial from the coupling constants ( $\delta$ 3.49, dddd,  $J=12.5, 12.5, 4.5, 4.5$  Hz). Therefore, the structure of **3** was identified as 5 $\beta$ -hydroxyilicic acid. This was finally confirmed by X-ray diffraction analysis (**Figure 3**).

Cytotoxicity tests were conducted on KB cells. All the three new compounds exhibited some cytotoxic effects with  $IC_{50} > 10^{-4}$   $\mu$ M.

**Figure 3** X-ray structures of compounds **1** and **3**



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7. <sup>13</sup>C-NMR spectral data of compounds **1-3**. (**1**): C-1 – C-15: 126.0, 201.9, 42.9, 37.3, 41.7, 30.0, 33.8, 30.3, 40.7, 146.3, 170.2, 177.6, 123.6, 19.3, 15.7; (**2**): C-1 – C-15: 37.1, 22.8, 124.9, 131.0, 143.2, 121.6, 38.5, 26.3, 38.2, 31.3, 145.5, 126.0, 172.6, 23.4, 20.1; (**3**): C-1 – C-15: 38.8, 18.2, 38.7, 76.5, 76.8, 36.7, 38.3, 27.6, 35.2, 38.8, 148.4, 171.2, 122.4, 25.6, 26.8. <sup>1</sup>H-NMR spectral data of compounds **1-3**. (**1**): 5.94 (br s, 1H, H-1); 2.34 m, H-3; 2.40 m, H-3'; 2.36 m, H-4; 2.41 m, H-6 $\alpha$ ; 1.98 m, H-6 $\beta$ ; 2.63 (dddd 1H  $J=11.0, 11.0, 4.5, 4.5$ Hz, H-7 $\alpha$ );

2.00 m, H-8; 2.76 (dddd 1H J=13.5, 11.0, 11.0, 4.5Hz, H-8' ); 1.70 (ddd 1H J=13.5, 11.0, 4.5Hz, H-9 $\alpha$ ); 1.92 (ddd 1H J=13.5, 4.5, 4.5Hz, H-9 $\beta$ ); 6.26 (br s, 1H H-13); 5.70 (br s, 1H H-13'); 1.16 s, H-14; 1.05 d (6.0), H-15. (2): 1.60 m, H-1 $\alpha$ ; 2.05 m, H-1 $\beta$ ; 2.06 ddd (12.5, 4.5, 4.5), H-2 $\alpha$ ; 2.64 ddd (12.5, 11.0, 4.5), H-2 $\beta$ ; 5.56 br s, H-3; 5.39 br s, H-6; 3.42 ddd (10.0, 7.5, 3.0), H-7 $\alpha$ ; 1.44 m, H-8 $\alpha$ ; 1.40 m, H-8 $\beta$ ; 1.54 m, H-9 $\alpha$ ; 1.56 m, H-9 $\beta$ ; 6.34 br s, H-13; 5.70 br s, H-13'; 1.00 s, H-14; 1.79 s, H-15. (3): 1.02 ddd (13.5, 4.8, 4.8), H-1 $\alpha$ ; 1.76 ddd (13.5, 11.0, 4.8), H-1 $\beta$ ; 1.90 (dddd 1H J=13.5, 12.0, 7.0, 4.5Hz, H-2 $\alpha$ ); 1.68 (dddd 1H J=13.5, 7.0, 4.5, 4.5Hz, H-2 $\beta$ ); 1.38 d(dd 1H J=13.5, 6.8, 4.5Hz); 2.68 (ddd 1H J=13.5, 12.0, 6.8Hz, H-3 $\beta$ ); 2.06 (dd 1H J=13.5, 4.5Hz, H-6 $\alpha$ ); 1.48 (dd J=13.5, 12.0Hz, H-6 $\beta$ ); 3.49 (dddd 1H J=12.0, 12.0, 4.5, 4.5Hz, H-7 $\alpha$ ); 1.18 m, H-8 $\alpha$ ; 1.72 m, H-8 $\beta$ ; 1.70 (ddd 1H J=13.2, 9.8, 3.5Hz, H-9 $\alpha$ ); 1.28 (ddd 1H, J=13.2, 3.5, 3.0Hz, H-9 $\beta$ ); 6.10 (br s, H-13); 5.56 (br s, H-13'); 0.99 s, H-14; 1.26 s, H-15.

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